



Chemical, Spectroscopical and Chemiometric Analyses of Vegetable Oils Submitted to Thermal Stress

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Abstract

In this work, chemical transformations and kinetic behavior of raw soybean oil and olive oil samples were evaluated by traditional and spectroscopic methods after continuous thermal stress. Initially, a multivariate classificatory study (PLS-DA) was carried out, using data from mid-infrared spectroscopy to differentiate 44 commercial oils samples (olive, canola, sunflower, corn and soybean). In this step, 19 important spectral regions were selected for samples classification. Two figures of merit were calculated (sensitivity and specificity) with this model presenting values above 83.3% of sensitivity for all classes and above 71.4% for 4 classes in specificity. In the next stage, soybean oil and olive oil samples were subjected to thermal stress for 15 h (5 h / day) at 200 °C. The results obtained by tradition methods as total acidity index demonstrated that soybean oil has a greater variation when compared to olive oil, and that after 4 and 11 hours of heating, soybean oil and olive oil, respectively, were outside the Brazilian Health Surveillance Agency standards. For the peroxide index, it was possible to determine that both oils suffered oxidation and the results demonstrated also that soybean oil suffered a greater degradation in relation to olive oil. Finally, a PCA, using mid-infrared data from the kinetic studies of soybean and olive oils thermal degradation, was carried out and demonstrated that molecular changes occurred basically in 8 spectral regions, resulting from the formation of sub products resulting from the thermal oxidation of the fatty acids presented in the samples.

Keywords: Vegetable oils; mid infrared; thermal stress; PLS-DA; PCA

Introduction

Vegetable oils represent one of the main products extracted from plants, and a large part of this production is destined for food products. These oils could also be used as raw materials for paints, varnishes, lubricants and cosmetics [1-3]. Although, when used as food, they are important sources of lipids [1-6]. However, it is known that foods containing oils and fats oxidize during storage at oxygen-presenting atmosphere, due to self-oxidation. In addition, when these foods are exposed to high temperatures (fried foods), this oxidation process becomes more accelerated, occurring oxipolymerization reactions and thermoxidative decomposition [7,8]. A series of complex reactions occurs during the frying process, causing irreversible changes in the structure of oils and fats. These changes are caused due to: (i) the moisture, which causes hydrolytic reactions of the triacylglycerol molecules (TG); (ii) oxygen from the air, which comes into contact with the food through the surface of the recipient, leading to oxidation reactions and, (iii) temperature, approximately 200 °C, which causes thermal changes [9-11]. It is common to consume foods

after frying processes with vegetable oils [12-15]. At the end of the process, certain foods submitted to this process have about 50% of total polar compounds formed, resulting from the oxidation of oil and fats (Brazilian Health Surveillance Agency - Anvisa, regulates that this value does not exceed 25%) [16]. These polar compounds can cause reactions on the human body, such as: irritations of the intestinal tract, diarrhea and reduced growth [17,14]. One of the greatest difficulties found in determining the maximum frying time that a vegetable oil can be subjected to, and remaining within the parameters described by ANVISA, occurs because they are generally used in different foods and under different conditions, which may influence the oxidation speed or not [16]. Due to the high consumption of these oils, new studies are always interested in evaluating the maintenance of physical-chemical, sensory and organoleptic properties of fried foods [18]. In this case, the present work proposes, at first, (i) a spectroscopic and chemometric study for evaluate the differences between five vegetable oils, followed by a (ii) second study of the chemical changes in the process of

frying olive and soybean oils discussing the main spectral regions affected by that. Finally, the work presents results (iii) of iodine and peroxide index obtained by classical methods of analysis.

Material and Methods

Reagents

The reagents used in this work presented analytical grade and belonged to the Chemistry Laboratory from Ifes - campus Vila Velha.

Samples

Forty-four refined soybean oils (*Glycine max*), corn (*Zea mays*), sunflower (*Helianthus annuus*), canola (*Brassica campestris*), and olive oil (*Olea europea*) were purchased from local markets in Vitória City, ES, Brazil.

Mid-Infrared spectroscopy (MIRS) analysis of the vegetable oils

The forty-four samples of five vegetable oil types were analyzed (olive oil, canola, sunflower, corn and soybean) by mid-infrared spectroscopy. Aliquots of 5 μl of each vegetable oil sample were added to commercial KBr tablets and analyzed. The IR spectra were collected on a Spectrum BX 79589 Perkin Elmer spectrometer, using 20 scans between 4000 - 400 cm^{-1} with 2 cm^{-1} resolution. All samples were collected in triplicate.

Vegetable Oils Thermal Stress

Approximately 1000 mL of soybean and olive oils samples were heated on a heating plate with controlled temperature at 200 $^{\circ}\text{C}$, for 3 days, 5 hours a day, in a refractory glass bottle (PIREX[®]). The aliquots of the analyzed oils were collected every hour and at the end of each heating day.

Free Fatty Acids Quantitative Determination (% of oleic acid)

To determine the amount of free fatty acids (FFA), 2 g of each oil sample were dissolved in 25 mL of isopropanol and 1 mL of phenolphthalein (0.1%). The solutions were titrated with standard solution of potassium hydroxide 0.1 $\text{mol}\cdot\text{L}^{-1}$, until the appearance of free pink color (% of oleic acid) [19, 20].

Peroxide index determination (PI)

For the determination of the peroxide index, 5 g of each aliquot of vegetable oil were dissolved in 30 mL of acetic acid/chloroform solution (3:2 v/v) and 0.5 mL of saturated potassium iodide solution (60 %). The solutions were kept at rest for one minute and then 30 mL of distilled water and 0.5 mL of starch solution [1%] were added. The mixtures were titrated with a standard solution of sodium thiosulfate 0.1 $\text{mol}\cdot\text{L}^{-1}$, until the bluish color disappeared. A blank test was performed under the same conditions described, without the presence of the sample [19,20].

Iodine Index Determination ($\text{mEq}\cdot\text{kg}^{-1}$)

For the iodine index determinations, aliquots of 0.25 g of each vegetable oil sample and 10 mL of cyclohexane were mixed in a 500

mL erlenmeyer flask with a lid. Then, 25 mL of Wijs' solution was added to the mixture, under mechanical stirring. After the addition, the solution was left to stand in the dark and at room temperature for 30 minutes and then 10 mL of potassium iodide solution 15% (W/V) and 100 mL of cold distilled water were added. Finally, the solution was titrated with a standard solution of 0.1 $\text{mol}\cdot\text{L}^{-1}$ sodium thiosulfate until a faint yellow color appeared. At that moment, 1 to 2 mL of 1% starch indicator solution was added and the titration was continued until the blue color completely disappeared. A blank determination was performed using the same method [21].

Chemometric Treatments

The original spectroscopic profiles were organized into a matrix format X ($I \times J$), where each replicate was considered one sample. Data analysis was carried out using Matlab v.2017 software (The MathWorks, Co., Natick, MA, USA) with the PLS_Toolbox computational package (Eigenvector Research, Inc. – PLS_Toolbox version 8.61.) [22]. Four pre-treatments were applied to the original data matrix to minimize signal/noisy ratio, baseline problems and different days of analysis: Savitzky- Golay smoothing with a window size of 11 points [23], first derivative, normalization and the data was mean centered. The Discriminant Analysis by Partial Least Squares (PLS-DA) was used for vegetable oils samples classification, while the Principal Component Analysis (PCA) was used in the kinetic degradation studies with soybean and olive oils samples [24]. Variable selection was carried out by the ordered predictors selection method (OPS) [25] and visual inspection.

Results and Discussion

Vegetable Oils Classificatory Analysis by PLS-DA

From the medium infrared spectra obtained by the five types of vegetable oils studied, it was possible to build a multivariate classification model known as PLS-DA. For this, the original data, X , was pre-treated and the important variables for discrimination of the oil classes were determined using the OPS algorithm [25] and visual inspection. Figure 1 shows the 200 selected variables (19 spectral regions) for the construction of the PLS-DA model, while Table 1 describes each spectral region in more detail. The PLS-DA model was built with 5 latent variables (VL) responsible for describing 98.17% of the data in block X , presenting a calibration error near 0% and an error between classes in the amount of 0.176%. According to the first latent variable (LV1) indicated in Figure 2, it is possible to observe the discrimination of 3 classes of vegetable oils (olive, canola and corn oils), in addition to the overlap of two other classes (soybean and sunflower oils). This overlapping of the soybean and sunflower classes occurs, once these oils have very close mono and polyunsaturated fatty acid compositions, causing the proposed model to present small flaws when classifying them. The loading analysis in LV1 (Figure 3) demonstrated that the spectral regions 1 (3025 - 3001 cm^{-1} - CH stretch in sp^2 carbons), 4 (2925-2919 cm^{-1} - CH stretch in sp^3 carbons) and 6 (1752 - 1746 cm^{-1} - stretches C = O) are the main responsible for the separation of olive oil and canola samples (negative loadings). The disposition

of the classes in LV1 (Figure 2) has a good association with the composition of mono and polyunsaturated fatty acids shown in the literature, with even overlapping in samples that have similar fatty acid profiles [16]. The olive oil samples were arranged on the negative side of LV1, as it is the vegetable oils (VO) that present the highest percentage of monounsaturated fatty acids, followed by canola, corn, sunflower and soy oil. Other spectral regions that present an interesting variability between the analyzed VOs are

the regions from 10 to 17 (positive loadings in Figure 3). In oils with a higher degree of establishment these regions become more intense than in oils with a lower degree, such as olive oil and canola [26]. From the construction of the PLS-DA model, it was possible to calculate two important figures of merit to demonstrate the validity of the data obtained. Therefore, the results for sensitivity and specificity are shown in Table 2.

Table 1: Selected wavenumbers for PLS-DA oils classification.

Region	Wavenumber (cm ⁻¹)	Spectral informations
1	3025 - 3001	alkenes C—H stretch
2	2993 - 2998	aliphatic C - H stretch (1 st , 2 nd e 3 rd)
3	2968 - 2948	aliphatic C - H stretch (CH ₃ , CH ₂ , first and second carbon.)
4	2925 - 2919	aliphatic C - H stretch (CH ₃ , CH ₂ , first and second carbon.)
5	2854 - 2852	aliphatic C - H stretch (CH, tert. carbon)
6	1752 - 1746	C = O stretch (carboxylic acids)
7	1457 - 1455	C - H (CH ₂) _n Deformation
8	1421 - 1419	C - H (CH ₂) _n adjacent deformation to carbonyl
9	1403 - 1397	C - O - H stretch
10	1393 - 1383	C - H deformation (t-butyl ou isopropyl)
11	1263 - 1260	C - O stretch for carboxylic acids and ethers
12	1157 - 1152	C-O stretch of esters, ethers, alcohols and phenols
13	1143 - 1136	C-O stretch of esters, ethers, alcohols and phenols
14	1127 - 1090	C-O stretch of esters, ethers, alcohols and phenols
15	1075 - 1071	C-O stretch of esters, ethers, alcohols and phenols
16	1036 - 1032	C-O stretch of esters, ethers, alcohols and phenols
17	1007 - 980	Angular deformation outside the plane RCH=CH ₂
18	964 - 961	Angular deformation outside the plane RCH=CH ₂
19	926 - 918	O-H, Broadband (angular deformation) of medium intensity, due to the angular deformation outside the plane of C = O from carboxylic acids.

Table 2: Classification results from the vegetable oils analyzed by mid-infrared spectroscopy.

Class	Corrects	False	False	Sensitivity(%)	Specificity (%)
		Positive	Negative		
Olive oil (1)	42/42	1	0	100	97.7
Soybean oil (2)	32/33	7	1	96.7	82
Canola (3)	21/21	1	0	100	95.5
Corn (4)	15/18	6	3	83.3	71.4
Sunflower (5)	15/18	19	3	83.3	44.1

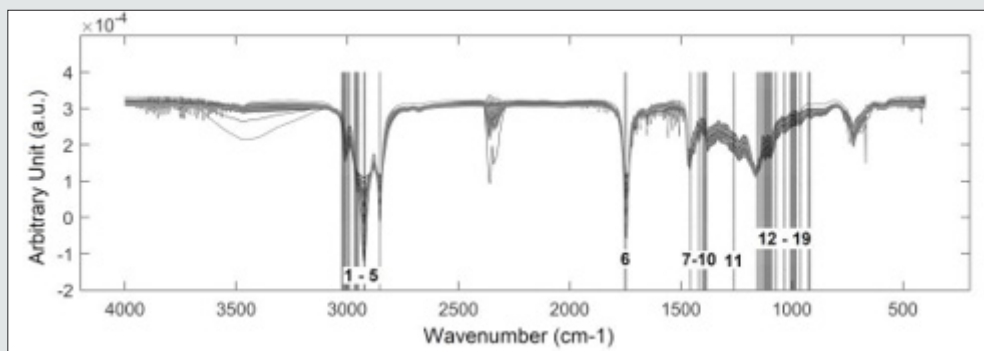


Figure 1: Selected variables for the PLS-DA model constructed.

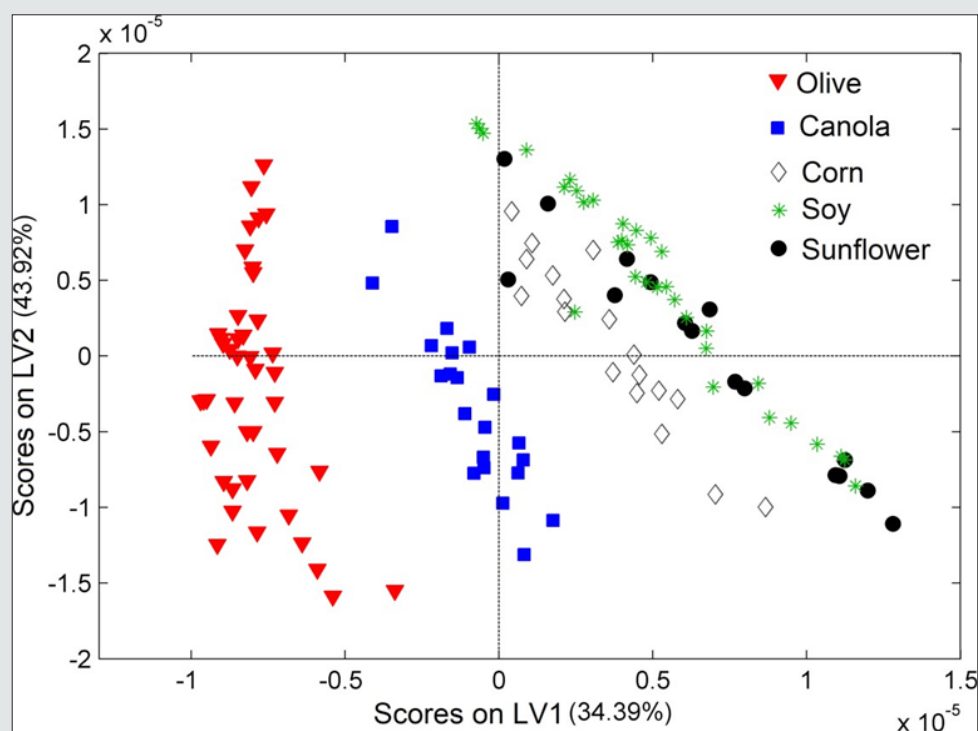


Figure 2: Graphical Scores between the latent variables 1 and 2.

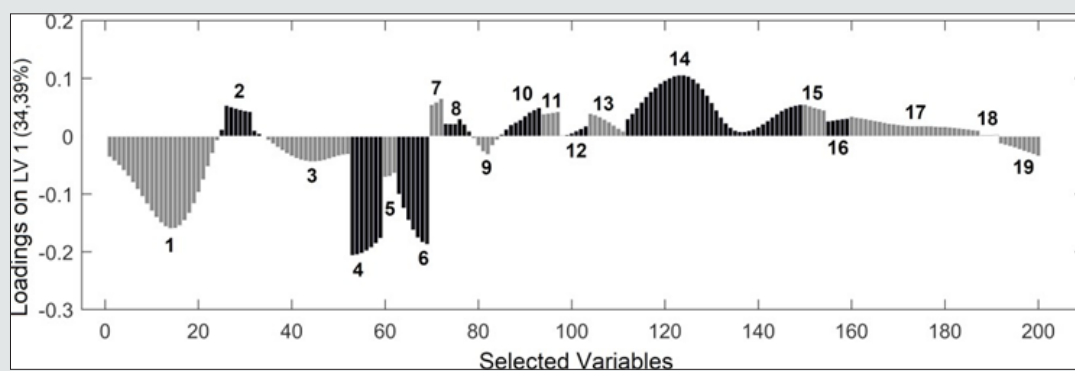


Figure 3: Loadings from selected variables for PLS-DA model construction

Vegetable Oils Thermal Stress

In the study on thermal stress, 2 types of vegetable oil (VO) were used (soybean and olive oils). These oils were chosen because they have different levels of mono and polyunsaturated fatty acids (FA) (results above - Figure 2). Thus, the samples were subjected to prolonged heating for 5h / day for 3 consecutive days at an average temperature of 200 °C. The choice of this temperature was due to a study found in the literature, where it is explained that commercial

oils have antioxidants in their composition, and these decrease their efficiency at temperatures above 180 °C [1]. The thermal stress of VO can lead to the formation of polymeric compounds when in an advanced oxidized state, causing changes in the viscosity and color of vegetable oils [26]. These changes were more evident in samples containing soybean oil, because in samples with a higher percentage of polyunsaturated fatty acids, there is a greater propensity to polymerization reactions (Diels-Alder reaction) and consequent darkening of the samples as shown in Figure 4.

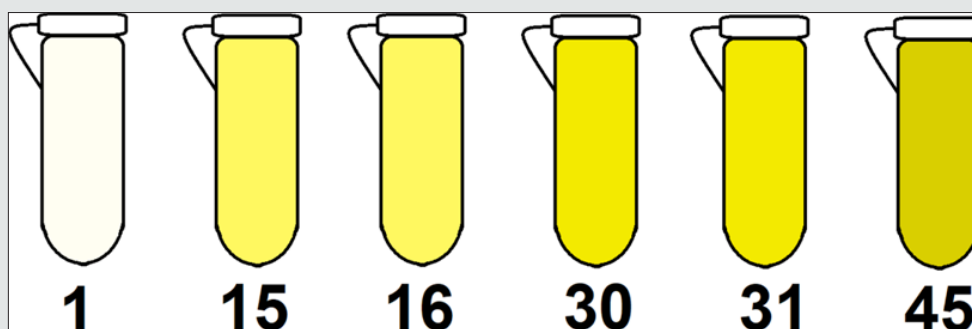


Figure 4: Schematic representation of color change in soybean oil samples at different heating times (1 to 45 h).

Exploratory Analysis of Soybean and Olive Oils Degradation by Thermal stress

Initially, the original spectra of the two oil samples were overlaid and pre-treated (mean centered, smoothed by Savitzky-Golay with 11-points window, derived and normalized). Subsequently, a visual selection of the variables was performed in order to determine spectral regions that showed systematic differences between the samples in 3 frying times (0h, 8h and 14h), as can be seen in Figure 5 (Soybean - A and Olive B) and Table 3. Two Principal component analyses were performed with the spectral regions selected for the oils. In the case of soybean oil sample, according to the PC1 (67.72% of variance) shown in Figure 6A, it is possible to observe 3 different groups of samples. These groups, from right to left, represent samples subjected to thermal stress for different times: with 0h (▼), 8h (■) e 14h of frying (●). On the other hands, in the PCA for olive oil (Figure 6B) the samples were almost completely displaced in PC1, but a small variation in PC2 can also be noticed (Accumulated variance in PC1 and PC2 - 88.73%). The loadings analysis in PC1 (Figure 7A) indicates a decrease in the band intensity

between 3008-2994 cm^{-1} related to the oxidation of double bonds and, consequently, formation of peroxides in degraded soybean oils. The variables selected in the overlapping spectra (4 regions) for soybean oil samples with long frying time indicate that there was a variation in the existing FAs in the samples. When analyzing region $n^{\circ} 1$, it can be noted that there was a decrease in the intensities of the bands in that region, indicating that there probably was a replacement of hydrogens in the α carbon of a double bond with a free radical, probably due to the autoxidation of FA. In region $n^{\circ} 2$ (also selected for olive oil), it can be seen that there were changes in the intensities and forms of the carbonyl group ($\text{C} = \text{O}$), suggesting the formation of new compounds such as: aldehydes, ketones or carboxylic acids, probably originated from the oxidation of hydroperoxides. Analyzing region $n^{\circ} 3$, it is possible to note the decrease in the bands in that region, probably indicating the degradation of trans-double bonds, which is a sign of the occurrence of oxidative processes [26]. It can also be noted that in region $n^{\circ} 6$ (also selected for olive oil) there was an increase in the bands, indicating the probable replacement of hydrogens by free radicals formed due to thermal degradation.

Table 3: Selected Wavenumbers for the two PCA.

Regions	Wavenumber (cm^{-1})	Spectral information's
1 ^a	3008 - 2994	Alkenes C—H stretch
2 ^{a,b}	1754 - 1730	C = O stretch (carboxylic acids)
3 ^a	1690 - 1674	C = C stretches (alkenes)
4 ^b	1551 - 1543	C = C stretch
5 ^b	1524 - 1509	C = C stretch
6 ^{a,b}	968 - 956	C - H deformation (R-CH=CH_2)
7 ^b	1190 - 1189	C - O stretch for carboxylic acids and ethers
8 ^b	1129 - 1128	C-O stretch of esters, ethers, alcohols and phenols

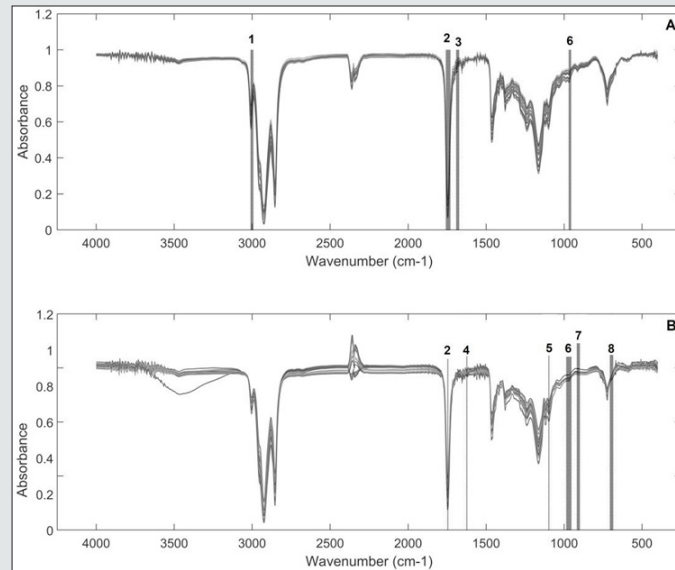


Figure 5: Overlapped spectra of soybean oil (A) and olive oil (B) samples.

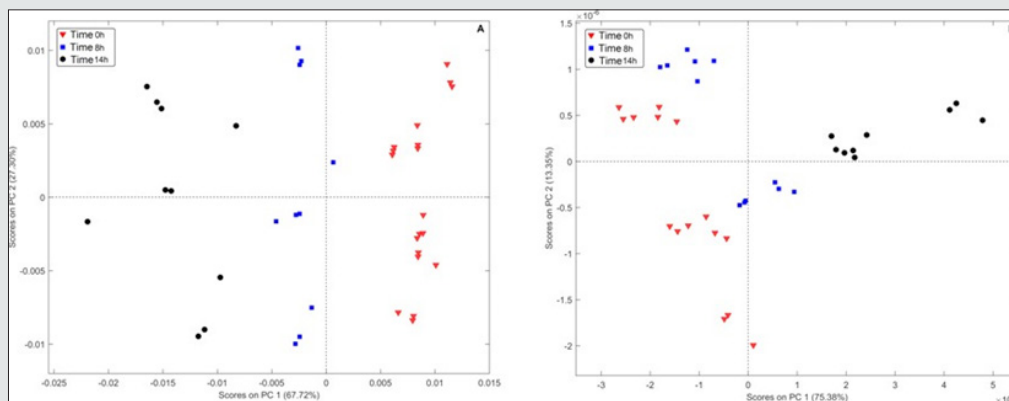


Figure 6: Graphical Scores of soybean oils samples between first and second PCs (A) and olive oils (B).

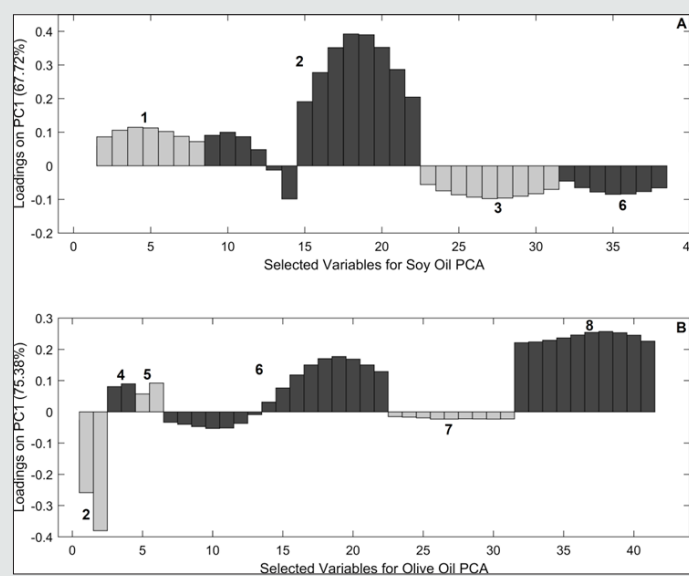


Figure 7: Loadings of selected variables in PC1 for Soybean oil (A) and Olive oil (B).

Classical Methods

Fatty acids, peroxide and iodine contents

The amount of free fatty acids (FFA) reveals the oxidation state of the oils and is defined by the amount (mg) of potassium hydroxide necessary to neutralize the free fatty acids present in 1g of oil or fat [27]. This measure is important, since the production of FFA is almost always associated with partial hydrolysis of glycerides, which occurs at a higher speed due to exposure to light or heating [28, 29]. According to Table 4, it can be seen the values found for the olive oil (OO) and soybean oil (SO) samples. The results obtained indicate that after 11 and 4 hours of heating, OO and SO, respectively, are outside the standards regulated by resolution N°. 482 of ANVISA (2005), which for the extra virgin OO is at most 1% and for the SO it is at most 0.3%. The consumption of these oils with a high total acidity index (free fatty acids) can lead to mild or even severe gastrointestinal problems depending on the amount consumed [17]. Thermal degradation causes changes in viscosity, odor and color, and these changes are due to oxidative processes and at that point, it is important to analyze the peroxide index [29]. This parameter is one of the most used tests to assess the degree of oxidation of oils. According to ANVISA, the maximum permitted value of peroxides for olive oil and soybean oil are 20 mEq / kg and 10 mEq / kg, respectively [16]. Samples with a high peroxide content indicate a high degree of deterioration [29]. According to Table 4, it can be found the peroxide index values for the samples of OO and SO. Also it can be noted that the soy oil becomes unfit for consumption after 3 hours of frying while the olive oil does not exceed the limit during the experimental time. These data demonstrate that monounsaturated oils (OO) are more suitable for consumption than polyunsaturated oils (SO). It is worth mentioning that the peroxide index indicates the formation of intermediate compounds such as peroxides and hydroperoxides (initial stage). Over time, these hydroperoxides break down into other, more stable compounds such as aldehydes, ketones and carboxylic acids [26].

In the two oils studied, the variation in its chemical and physical properties was noticeable, and from the analysis of the peroxide index, it was evident that there was an effective oxidation of the oils. It was also possible to notice, that over time, the concentration of peroxides begins to decline. a sign that oxidation is already in a new stage, it can be the stages of propagation or termination. Lastly, the iodine index is a qualitative parameter that can be used to predict the presence of insaturation in samples of vegetable oils. This index is very useful for determining the degree of installation of the samples and for being able to serve as a comparison with the peroxide index, once, they are tests that must present opposite results [28,29].

In that parameter, the higher the value found, the greater the degree of insaturation of the sample in question [30]. The results for the iodine index are also shown in Table 4. Through the results it is evident the occurrence of the break of insaturation present in samples of SO as well as in samples of OO. So that a deeper comparison between the results obtained with the oils studied was possible, the data in Table 4 were normalized and transformed into a graph (Figure 8). According to Figure 8, it can be seen that the relative acidity present in soybean oil. at the end of the process. it is greater than the acidity found in olive oil. This result may appear due to the secondary products of thermal degradation (aldehydes, ketones and carboxylic acids). since the OS for having a greater degree of establishment than the Az. will present a greater formation of these compounds. Another fact to highlight is the relationship between the iodine (II) and peroxide (IP) indices, as both are classic tests for assessing the oxidation status of oils, however, samples with high levels of iodine content should have low levels of peroxides and so on, successively. Fact confirmed by Figure 8. This opposite relationship occurs, because the PI indicates the degree of oxidation of the VO, since reactions occur in the insaturation of the FA molecules and the PI is a test that qualitatively predicts the presence of the insaturation [31].

Table 4: Free fatty acid, peroxide and iodine contents for olive oil and soy oil, subjected to heating for different periods.

Time	Total acidity - OO (%)	Total acidity - SO (%)	Peroxides - OO (mEq/kg)	Peroxides - SO (mEq/kg)	Iodine - OO	Iodine - SO
0h	0.30 ± 0.00	0.10 ± 0.00	9.70 ± 0.10	7.20 ± 0.90	91.40 ± 0.30	122.80 ± 0.70
1h	0.30 ± 0.10	0.20 ± 0.10	9.80 ± 0.10	7.80 ± 0.00	91.30 ± 0.20	122.20 ± 0.20
2h	0.40 ± 0.00	0.20 ± 0.10	9.80 ± 0.10	8.40 ± 0.90	90.90 ± 0.20	121.10 ± 0.30
3h	0.40 ± 0.10	0.30 ± 0.00	10.30 ± 0.90	9.70 ± 0.10	89.90 ± 0.40	120.00 ± 0.40
4h	0.50 ± 0.00	0.30 ± 0.10	11.60 ± 0.10	11.10 ± 1.00	88.50 ± 0.20	119.10 ± 0.50
5h	0.60 ± 0.10	0.40 ± 0.00	11.60 ± 0.00	12.30 ± 0.90	87.00 ± 0.10	116.50 ± 0.30
6h	0.70 ± 0.10	0.50 ± 0.10	11.70 ± 0.10	13.10 ± 1.00	86.40 ± 0.00	115.80 ± 0.30
7h	0.70 ± 0.10	0.50 ± 0.10	12.70 ± 1.00	14.90 ± 0.90	85.40 ± 0.30	111.40 ± 0.70

8h	0.80 ± 0.02	0.50 ± 0.01	13.50 ± 0.10	15.60 ± 0.00	85.30 ± 0.30	106.30 ± 0.80
9h	0.80 ± 0.10	0.60 ± 0.10	14.70 ± 0.90	17.40 ± 0.20	85.10 ± 0.40	105.40 ± 0.20
10h	0.90 ± 0.00	0.70 ± 0.00	16.60 ± 0.90	13.60 ± 0.10	85.30 ± 0.50	102.40 ± 0.30
11h	1.00 ± 0.10	0.70 ± 0.00	16.80 ± 0.90	11.60 ± 0.90	85.10 ± 0.20	102.40 ± 0.30
12h	1.00 ± 0.00	0.80 ± 0.00	13.40 ± 0.10	11.10 ± 0.90	84.30 ± 0.30	100.50 ± 0.30
13h	1.20 ± 0.10	0.90 ± 0.10	11.50 ± 0.10	10.30 ± 0.90	83.30 ± 0.30	99.10 ± 1.00
14h	1.30 ± 0.00	0.90 ± 0.00	10.80 ± 0.90	9.00 ± 0.90	83.20 ± 0.20	95.30 ± 0.50

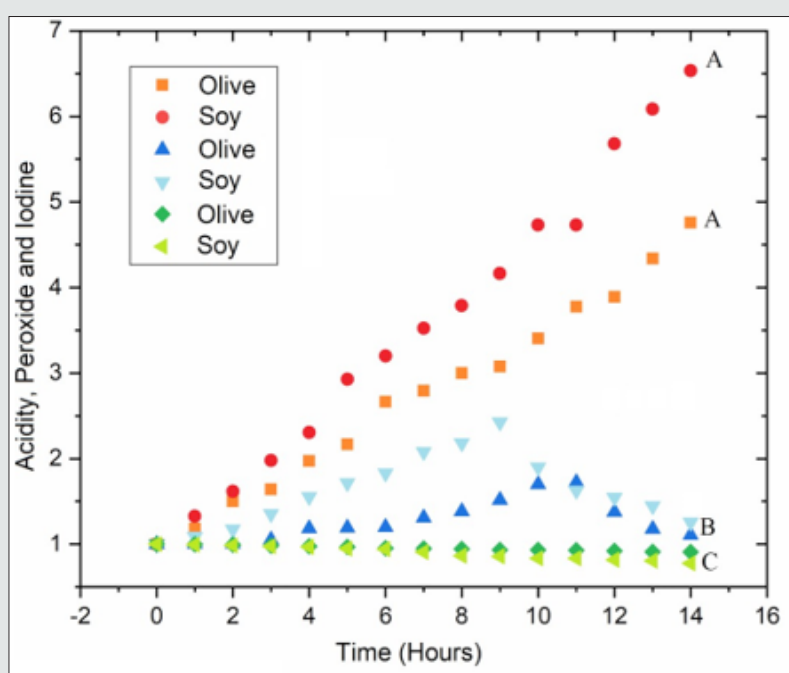


Figure 8: Table 4 data normalized and shown graphically. Total acidity (A). Peroxides (B) and iodine (C).

Conclusion

In this present work, it was possible to investigate the changes in vegetable oils, when subjected to thermal stress. This investigation took place through analytical quality verification tests regulated by ANVISA and also by spectroscopy in the mid infrared region and chemometrics. Thus, when performing the classificatory analysis of the vegetable oils most used commercially (olive oil, Canola, Sunflower, Corn and soybean oli), it was possible to find the main bands that can be used to separate these oils (19 spectral regions). By obtaining the main regions that differentiated commercial oils, it was possible to build a satisfactory classification model for predicting new samples, since 3 types of oils (olive oil, canola and corn) are easily separated while the other 2 types are overlapping

(soy and sunflower) due to the similarity in the mono and polyunsaturated fatty acid compositions. When the samples were subjected to thermal stress for 45 h at 200 °C (on average), it was possible to detect the acidity index and iodine and peroxides values. In both cases, the samples showed higher levels than the tolerated ones by the Brazilian legislation at the end of the process, making its consumption inappropriate after 4 hours of heating for soybean oil and 11 hours for olive oil. With the same samples subjected to thermal stress it was possible to make an exploratory kinetic study of the degradation through the spectra obtained in the mid-infrared confirming the results of the classic tests obtained and proving that during prolonged heating and at high temperatures, vegetable oils become more susceptible to thermal oxidation.

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