



Antibacterial Effects of Carotenoid Pigment Extracted from *Rhodotorula Glutinis* Strains on *Staphylococcus Aureus* Isolated from Mastitis Samples of Dairy Cows

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Abstract

Background: *Staphylococcus aureus* is the most common cause of bovine mastitis, which is often difficult to cure and leads to a lot of economic losses. Therefore, new antibacterial compounds (Especially natural compounds) are being investigated as an alternative to treat infections caused by these bacteria.

Objective: The aim of this study was to investigate the effect of pigments extracted from *Rhodotorula* against planktonic growth of *S. aureus* isolates.

Methods & Materials: In this descriptive research study, 100 milk samples were taken from clinical mastitis cow and isolation of *S. aureus* and *Rhodotorula* yeast was performed using standard microbiological tests. Extraction of carotenoid pigment of *Rhodotorula* yeast used to identify antimicrobial effect against *S. aureus* isolates and was measured by micro-dilution test. Scanning electron microscope (SEM) was also used to confirm the effect of the pigment on *S. aureus* isolates.

Results: A total of 25% of milk samples were infected with *S. aureus*, which confirmed by molecular method. A species of *Rhodotorula* was also isolated in one case which was confirmed as *R. glutinis* by PCR method (identification of its gene). The Geometric mean of MIC (Minimum Inhibitory Concentration) and MBC (Minimum Bactericidal Concentration) values of the carotenoid pigment was 39.36 and 78.82 $\mu\text{l.ml}^{-1}$, respectively.

Conclusions: In general, we showed that carotenoid pigment from *R. glutinis* can be effective agents planktonic form of *S. aureus* isolates. Therefore, it can be claimed that the carotenoid pigment *Rhodotorula* is a good candidate for controlling *Staphylococcus* infections. However, many studies and clinical trials should have been performed on it.

Keywords: Mastitis; *Staphylococcus aureus*; *Rhodotorula glutinis*; Pigment; SEM

Introduction

Mastitis is one of the most common and economically important diseases in dairy cows in the world, so it causes a lot of damage

to the livestock industry every year. It considerably affects milk production, animal welfare, and food safety [1]. More than 140 differ-

ent pathogenic species have been reported in scientific papers as factors in mastitis [2]. The majority of mastitis cases are produced by a relatively small group of bacteria, including *Staphylococcus aureus*, *Streptococcus aureus*, *Mycoplasma spp* and coliforms [1,3].

S. aureus is one of the most important and common bacteria in the clinical development and subclinical mastitis [4,5]. It is also considered as one of the incriminated pathogenic in humans and causes numerous skin and systemic diseases. Beta-lactams are the most successful group of antibiotics ever used to treat mastitis infections caused by *S. aureus* isolate. Unfortunately, resistance toward all B-lactam antibiotics has eventually been observed [6-8], therefore, new complementary treatment strategies (Nanoparticles, photodynamic therapy, herbal compounds, microbial pigments and ...) seem to be effective against resistant pathogens [9]. Among antimicrobial compounds, bacterial pigments are great alternatives to antibiotics because resistance toward them has not been reported among microorganisms, therefore it has attracted the attention of many researchers [10]. The pigments produced by microorganisms includes monascins, violacein, indigo, melanin, flavins, quinones, and more specifically carotenoids, showed distinct antibacterial effect against many pathogenic bacteria [11]. Numerous microorganisms have the ability to produce pigments. However, actinobacteria, rhodococcus and a number of fungi and yeasts including *Rhodotorula*, have the highest ability to produce pigments [12,13].

Rhodotorula glutinis is one of the most important and useful fungi in the production of various pigments [12]. This yeast has been used industrially in the production of carotenoid pigments and as a biological control agent for post-harvest diseases of fruits. Carotenoids belong to the chemical group of isoprenoid and fat-soluble polymers [14]. This pigment acts as a light-absorbing chromophore and produces yellow, red and orange colors [14,15]. Therapeutic effects of carotenoids include their use in the treatment of cancer, the treatment of cataracts and the treatment of cardiovascular diseases. However, few studies on the antibacterial properties of

this pigment have been done sparsely in the world [15]. Therefore, in this study, we tried to measure the antimicrobial effect of *Rhodotorula* pigment on *S. aureus* isolates, so that if there is a sufficient antimicrobial effect, more research can be done in this regard.

Materials and Method

Sample collection and yeast isolation

To isolate *Rhodotorula Spp*, different sediment samples (including soil, sludge, plants and flowers) were collected from around the Tehran University yards. From each sample 100 g were placed in plastic bag with an ice bag and transferred to microbiology laboratory in Tehran Veterinary college and then processed immediately to isolate yeast. Ten g of each sample was homogenized in 90 ml of 9% sterile saline, then cultured on yeast-peptone-dextrose (YPD) agar extract. All plates were incubated at 28 °C for 24 hours to grow the yeast colony and determine its morphology [12].

Molecular identification of yeast isolates

Molecular method was used to confirm the species suspected of *Rhodotorula*. To do this, First, DNA of isolates was extracted using phenol-chloroform method, then a fragment sequence from ribosomal genome (ITS) was amplified and identified using a specific primer. Primer sequence and PCR product length are given in Table 1. PCR reaction was performed in a final volume of 25 µl including 12.5 µl of Mastermix (Amplicon, 2X), 0.5 µl of each primer (10 picomoles), 3 µl of template DNA and 8 µl of deionized water. The PCR amplification was performed under the following conditions: initial denaturation at 95°C for 8 min followed by denaturation at 95°C for 1 min, annealing at 58°C for 45 sec and extension at 72°C for 1 min (35 cycles) and a final extension at 72°C for 10 min. Finally, the PCR product was run on 1% agarose gel for 45 minutes [16]. The *sti* gene of *Rhodotorula* species was also sequenced using the Sanger method.

Table 1: Primers used in PCR assays.

Gene	Primer	Sequence 5'- 3'	Product size (bp)
<i>its</i>	ITS -forward	TCCGTAGGTGAACCTGCGG	550
	ITS -reverse	TCCTCCGCTTATTGATATGC	
<i>femA</i>	FEM -forward	AAAAAAGCACATAACAAGCG	132
	FEM -reverse	GATAAAGAAGAAACCAGCAG	

Sample Collection and *S. aureus* isolation

In total, 100 samples of cow's milk suspected of mastitis were taken from industrial farms in Famenin city in Hamedan province of Iran, over a period of three months and transferred to a microbiology laboratory. Milk samples were centrifuged at 4 °C at 8000 rpm for 10 minutes and the sediment was cultured using swap in blood agar medium. After 24 hours of incubation, *S. aureus* colonies were identified using standard biochemical tests such as catalase,

nitrate, and DNase and coagulase test [17,18].

Molecular identification of *S. aureus* isolates

For molecular confirming of *S. aureus* isolates, *S. aureus* species-specific primer was used. First the DNA of bacteria was extracted by boiling method, then primers designed by Mehrota were used to amplify the *femA* gene (Tables 1) [19]. PCR reactions and temperature program were performed as in section 1-1, only the annealing temperature was 58 °C.

Purification of the carotenoid pigment

First, mass cultivation of *Rhodotorula* yeast was carried out in MMS medium and placed in a shaker incubator at 25 °C, 200 rpm for 86 hours. The yeast cells were then centrifuged at 5000 rpm for 20 minutes and then were washed. To the obtained biomass was added a Hydrochloric Acid, 1.00 Normal, and transferred to a bain-marie at 70°C for 90 minutes. After acid removal by washing, the cells were immersed in a solution of acetone and methanol (1:1) for 24 hours. Eventually, the crude pigment was collected and concentrated by evaporation. Quantity of crude pigment was measured by adding the pigment into the dried 25 ml preweighed beaker. After evaporation of the solvent, the weight of crude pigment was measured and stored in sterile vial.

Evaluation of antimicrobial effect of *Rhodotorula* pigment

The Minimum Inhibition Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of ethanolic extract of pigment were evaluated by well micro-titer plate method according to the guidelines of CLSI 2018. Briefly, 100 µL of the bacterial inoculate (corresponding to 0.5 of the McFarland) and 100 µL of the two-fold serial dilutions of the pigment (0.25 to 256 µL. mL⁻¹) were distributed into each well of ELISA microplates (96 well). Dilution was done till tenth. Column eleventh, as a negative control, contained dimethyl sulfoxide (DMSO) and bacteria, and column twelfth, as positive control, contained DMSO, bacteria and Penicillin (1 mg/mL). After 24 hours of incubation of microplates at 37 °C, microtiter plates were visually scored. MIC is the minimum concentration at which microbial growth is inhibited. For determining the MBC, 0.1 ml of culture medium from each well was sub-cultured on Muller

Hinton agar plate. The MBC was considered as the lowest concentration of the pigment associated with no bacterial culture.

Electron microscope

Electron microscopy was used to evaluate the effect of carotenoid pigment of *Rhodotorula glutinis* on *S. aureus* isolates. For this purpose, bacteria were cultured in TSB medium and after adding carotenoid pigment to the medium (final concentration of carotenoid pigment equal to MIC / 2), a sterile lamellar with a size of 0.5 cm² was placed in the medium. After incubation of the samples for 24 hours at 37 °C, the lamellae were fixed with 2.5% Glutaraldehyde solution. The samples were then dehydrated with ethanol and dried at room temperature. The fixed bacterial samples were mixed with the desired nanoparticles and a 9 nm gold coating was applied to the samples by spattering device. Eventually, the samples were photographed by FESEM model TESCAN mira3 (made in the Czech Republic) in different magnifications with a voltage of 15 kV [20].

Results

Yeast isolation

In order to identify and isolate the *Rhodotorula* spp. colony, the samples were cultured in Potato dextrose agar and incubated for 48h at 37 °C (Figure 1). Then the colonies were examined macroscopically for size, shape, color, and margin. In total, one species (1/100) of *Rhodotorula* was isolated from mastitis samples which was identified as *R. glutinis* using physical characteristics and biochemical tests. Also, in molecular confirmation, *its* gene was amplified, and the desired band (550 bp) was observed in electrophoresis of PCR product (Figure 2).



Figure 1: *R. glutinis* colony in the cultivation Potato dextrose agar.

Isolation of *S. aureus*

A total of 25% (25/100) of milk samples contained *S. aureus* strain in biochemical tests. Also, the Molecular assay were also

completely compatible with biochemical tests and in search of *femA* gene using PCR test, all isolates carried this gene and in PCR electrophoresis, showed a band of 132 bp (Figure 2).



Figure 2: PCR products obtained from the amplification of *femA* and its genes. Lane L: a 100 bp DNA ladder; lanes 1: Positive controls (*S. aureus* ATCC 25923, *R. glutinis*), lanes 2: Isolates, lanes 3: Negative controls.

MIC and MBC

Broth microdilution test was used to determine the minimum inhibitor concentrate of pigment on *S. aureus* isolates and the results are shown in Figure 3. The MIC and MBC of pigment on *S. au-*

reus isolates from mastitis were between 2 - 128 µg / ml. The Geometric mean of MIC and MBC) values of the pigment on *S. aureus* isolates was 39.36 and 78.72 µl.ml⁻¹. The results of microdilution test showed that all isolates were susceptible to Rhodotorula pigment.

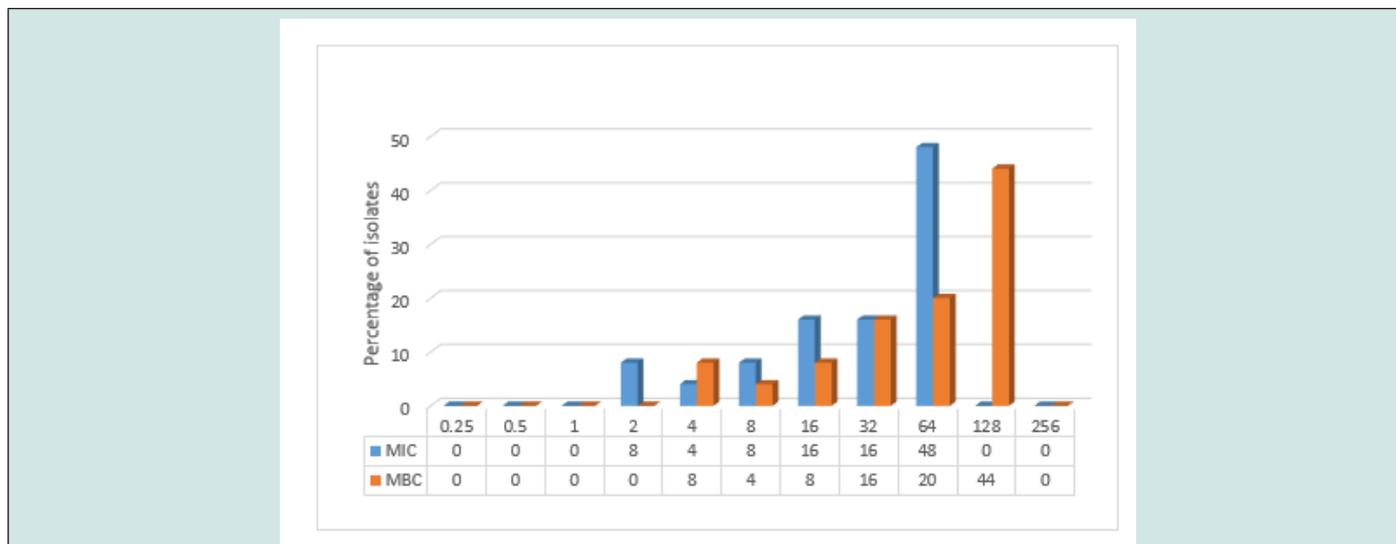


Figure 3: Broth microdilution test results.

SEM

SEM was employed to determine the inhibitory effects of the carotenoid pigment on *S. aureus* isolates. The results of SEM analysis showed that the carotenoid pigment at the sub-MIC concentration had a destructive effect on *S. aureus* isolates. As shown in Figure 4 -

Part A, *S. aureus* is placed on a glass slide in the form of a grape cluster. In this case, *S. aureus* is undamaged and has a healthy spherical shape. Figure 4-Part B & C, shows that the pigment destroys the bacterial cell wall by creating pores. In Figure 4-Part D, the bacterium *S. aureus* is completely destroyed.

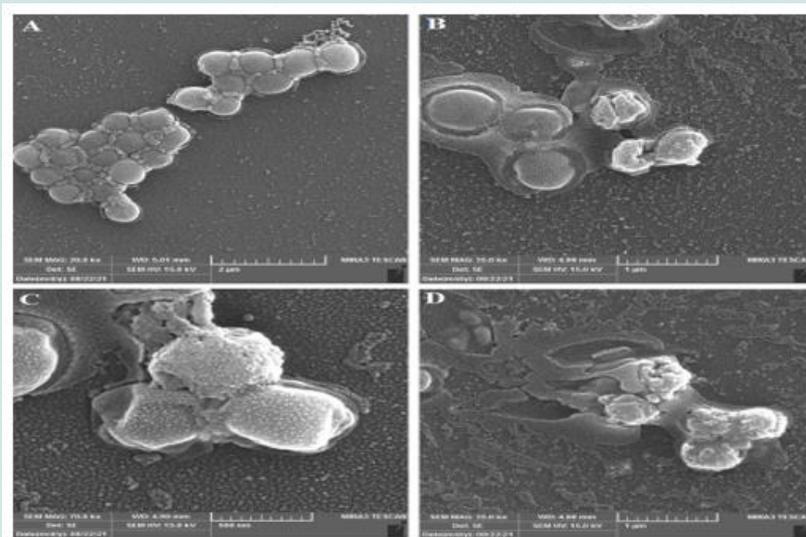


Figure 4: SEM results. Part A: Grape-like clusters of *S. aureus*, Part B, C and D: Destruction of the cell wall of *S. aureus* due to the application of carotenoid pigment of *R. glutinis*.

Discussion

Bovine mastitis is an inflammatory response of the udder tissue in the mammary gland caused due to physical trauma or microorganism infections [21,22]. It is one of the most important diseases of dairy cows, which leads to economic loss in dairy industries due to the reduction in the quantity and quality of milk and the reduction of the production life of infected cows. *S. aureus* has been the most predominant bacteria causing both agents of clinical and sub-clinical mastitis [22,23]. In the current study, a total of 100 milk samples were analyzed by biochemical and molecular test and results indicated that 25% were found positive for *S. aureus* isolation. Lower frequency of mastitis-associated *S. aureus* has been reported from maku, Iran (8%) [24], Northeast of Iran (10.3%) [25], Japan (7.7%) [26] and Argentina (20%) [27], in contrast with the higher frequency informed from Daland, Iran (20%) [28], Shiraz, Iran (31%) [29], Kurdistan, Iran (33.5%) [30], Bangladesh (72%) [31], Netherlands (42) [4], and India (53.3%) [7]. In general, the results of studies conducted in Iran and the world indicate the importance of *S. aureus* in causing mastitis in farms.

On the other hand, due to the increasing of drug resistance among *S. aureus* isolates, physicians and veterinarians have encountered difficulties in treating infections caused by this pathogen. Therefore, many researchers have tried to find new methods and antimicrobial compounds to control and prevent this problem [9,32]. One of the new, vital and bioactive compounds for controlling bacterial infections is the use of pigments produced by microorganisms [33]. Pigments such as carotenoids, melanins, violacein, indigo, monascins, flavins and quinones have been reported as good antimicrobial agents [34]. *Rhodotorula* yeast is a pigment-producing fungus. The genus *Rhodotorula* is abundant in nature and can be isolated from various sources such as air, seawater, plants, dairy products and the environment [35]. In a recent study, a species of

Rhodotorula was isolated from milk samples taken from cows with suspected subclinical infections, which was confirmed as a species of *R. glutinis* in molecular studies (Identification of *its* gene). The pigment obtained from *R. glutinis* yeast was purified and prepared, and its antimicrobial effect was measured on *S. aureus* isolates using broth microdilution method. According to the results of the microdilution broth test, the pigment extracted from *Rhodotorula* had an inhibitory effect on the growth of all isolates of *S. aureus* and at a concentration of 80 μ /ml, was able to kill *Staphylococcus* isolates.

Research have shown that the antimicrobial activity of carotenoids on bacteria have not been the same. The carotenoids produced by *Halomonas* have been reported to have antimicrobial activity against the antibiotic resistant *Klebsiella* sp., *Streptococcus pyogenes*, *Pseudomonas aeruginosa*, *Escherichia coli* and *S. aureus* [36]. In contrast, carotenoids extracted from *M. luteus* and *M. roseus* did not (didn't) have a lethal effect on *Escherichia coli* but inhibited the growth of *Enterococcus faecalis* and *S. aureus*. Other studies have shown that the carotenoid pigment extracted from *Corynebacterium* sp., *Bacillus* sp., *Kocuria roseus*, and *Brevibacterium* sp. had good antimicrobial effects against *S. zillusus*, *S. aureus* ATCC 25923 (MSSA) and *Bacillus masserans* [37].

Since gram- negative bacteria had shown high resistance to antimicrobial agents against gram- positive bacteria, it is expected that the MIC of *Rhodotorula* pigment against gram-negative bacteria is more than 80 μ /ml. In this case, Yolmeh et al (2016) reported that gram-negative bacteria were highly resistant to *R. glutinis* pigment than gram- positive bacteria [38]. In another study, Yolmeh et al (2018) Showed that the carcinoid pigment *Micrococcus roseacea* had a stronger destructive effect against gram-positives than gram-negatives bacteria [39]. In general, it could be said that the type of bacteria and the presence of resistance genes, including efflux pumps, which are generally resistant to many antimicrobial

agents, can affect the MIC of pigments, however no specific resistance to pigments has been reported so far [40]. Using SEM is the most commonly method to study the effects of antimicrobial compounds on microbial structure [41]. In this study, SEM was also used to confirm the effect of carotenoid pigments produced by *R. glutinis* on *S. aureus* isolates. As presented in the figure 4, untreated bacterial cells had a normal shape and were arranged in clusters form, but these structures were rarely seen in pigment-treated *S. aureus* cells. In the treatment group (containing carotenoid pigment), *S. aureus* cells were deformed and a rupture in the cell wall and leakage of cytoplasmic contents were observed. The mechanism impact of carotenoid pigments produced by *R. glutinis* on bacteria has not been completely clear. Although researchers have shown that, natural compounds such as plant essential oils and pigments of microorganisms, nonspecifically destroy the bacterial cell wall and kill it.

Conclusion

In the present study, the prevalence of *S. aureus* in mammary gland quarters of dairy cows in traditional farms of Famenin city was very high, hence, it can be said that the outbreak of *S. aureus* leads to the economic losses for traditional farms. Therefore, it is recommended to prevent the spread of *S. aureus* among other cows as soon as possible. The results of antimicrobial activity of carotenoid pigments produced by *Rhodotorula* also showed that this substance at a concentration of 80 micrograms has a good antimicrobial effect against *S. aureus* isolates and can be considered a great candidate for a novel complementary treatment of MRSA infections in the future.

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